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# **Green synthesis and characterization of bisphosphonate conjugated gold nanoparticle with *cissus quadrangularis* extract to enhance orthodontic anchorage**

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**Abstract**--Bisphosphonates are increasingly being used to treat several skeletal disorders such as osteoporosis. Several studies have shown that localized bisphosphonates improve orthodontic anchorage. Adverse reactions of the drug can be further avoided through more targeted drug delivery of this drug. The use of gold nanoparticles in bone tissue regeneration has shown promising results. The combination of bisphosphonate and gold nanoparticles has been shown to produce more targeted action through systemic administration in osteoporosis. The Adverse Effects of the Gold nanoparticle such as toxicity can be reduced through green synthesis. *Cissus Quadrangularis* is a green edible medicinal plant used in Ayurvedic preparations to treat osteoporosis, general inflammatory conditions, ulcers, menstrual disorders, hemorrhoids, and a variety of other ailments. The purpose of this study was to green synthesize Bisphosphonate conjugated gold nanoparticles with *Cissus quadrangularis* extract and test their antimicrobial and cytotoxic activity. Green synthesis of Bisphosphonate conjugated gold nanoparticles using *cissus quadrangularis* extract was achieved and is characterised using visual colour change, uv spectrophotometer,

TEM analysis. Gold nanoparticles have good antibacterial activity against *Streptococcus mutans* (150 ug/ml – 17 mm zone of Inhibition) & *Lactobacillus* (150 ug/ml – 20 mm zone of Inhibition) and showed 20% lethality at 50 ul concentration.

**Keywords**---Bisphosphonate, Green synthesis, Gold nanoparticles.

## Introduction

Nanotechnology aims to design, create and control matter in the dimensional range of 1-100nm[1]. The use of materials in these dimensions provides an opportunity to modify various properties such as solubility, diffusivity, blood circulation half-life, drug release characteristics, and immunogenicity at the level of atomic or biomolecules[2,3]. Biological membranes in the human body are selectively permeable to Nanoparticles. The nanoparticles can be surface modified to attach a variety of ligands which enable them to be biosensors, molecular-scale fluorescent tags, imaging agents, and targeted drug delivery vehicles [4–6].

Nanoparticle synthesis by the top-down and bottom-up methods have been described in the literature [7–9]. In general, both methods of nanoparticles use high radiation or concentrated reductants and stabilizing agents that are harmful both to the environment and to human health. Green synthesis of nanoparticles is a type of bottom-up approach in which reaction occurs with the help of reducing and stabilizing agents which eliminates the use of expensive chemicals, consumes less energy, and generates environmentally benign products and by-products. They have also shown to reduce the toxicity of the nanoparticles. Hence nanoparticles synthesized by green synthesis are considered far more superior to those manufactured through other methods [10–13].

Bisphosphonates are increasingly being used to treat a wide range of skeletal tissues, including hereditary skeletal disorders in children, postmenopausal and glucocorticoid-induced osteoporosis (GIO), and cancer-related bone metastases, osteoporosis (juvenile, postmenopausal or involutional [senile], glucocorticoid-induced, transplant-induced, immobility-induced, and androgen-deprivation-related), Paget disease of bone, osteogenesis imperfecta (OI), hypercalcemia, and metastatic malignancy. Bisphosphonates can provide significant clinical benefit in diseases characterized by an imbalance between osteoblast-mediated bone formation and osteoclast-mediated bone resorption by destroying the osteoclasts by promoting apoptosis. As a result, bisphosphonates have emerged as an effective treatment for osteoclast-mediated bone resorption. Localized Bisphosphonates have been shown to enhance orthodontic anchorage in several studies[14–16].

Various nanoparticles have shown promise in the regeneration of bone tissue. Gold nanoparticles (GNPs) are a popular choice among these nanoparticles because they are effective at both promoting osteo-differentiation and inhibiting the formation of osteoclasts. GNPs have several advantages, including the ability to accelerate osteoblast differentiation, inhibit adipose-derived stem cell differentiation, suppress osteoclast formation, and promote bone formation in

bone tissue regeneration. GNPs have been reported to significantly increase bone tissue regeneration in rabbits. When injected into the body, however, GNPs can cause toxicity. As a result, the surface of these particles must be modified to specifically target bone tissue [17–20].

*Cissus quadrangularis* L. is a member of the Vitaceae family, and its common name is Perandai (Tamil). It has been used in Indian folk medicine to treat a variety of pathological conditions such as osteoporosis, general inflammatory conditions, ulcers, menstrual disorders, haemorrhoids, and many others [21]. The plant extract's ability to heal bones has been demonstrated. The phytochemicals present in the plant have been attributed with medicinal properties, and some of the main constituents are ascorbic acid, flavanoids such as quercetin, kaempferol, and luteolin, stilbenes such as resveratrol, quadrangularin A, and pallidol, terpenoids, gallic acid derivatives, and glucosides such as bergenin. These are known to have good antioxidant and reducing properties, which is why the stem extract of this plant was chosen for AuNP preparation. *Cissus quadrangularis* extracts have previously been used to produce biogenic nanoparticles such as silver, copper oxide, ferric oxide, palladium, and other metals. These nanoparticles have been shown to have antibacterial and cytotoxic properties against a variety of bacteria and cancer cells [22–28]. Bisphosphonate conjugated with gold nanoparticles has been shown to provide a more targeted action [29]. The goal of this study was to green synthesize Bisphosphonate conjugated gold nanoparticles with *cissus quadrangularis* extract and to study their antimicrobial and cytotoxic properties.

## **Materials and Method**

### **Green synthesis gold nanoparticles and conjugation of bisphosphonate**

Stem extract of *cissus quadrangularis* were dried in an oven at 30 °C and ground to a coarse powder. 1gm of available *cissus quadrangularis* stem powder was boiled at 100 degrees c with 100 ml of distilled water in a beaker. The extract was then filtered using a filter paper to obtain 75ml. To reduce the Au<sup>2+</sup> ions, 30 mL of the extract was added to a reaction vessel containing 70 mL of chloroauric acid solution. The nanoparticles were then centrifuged at 1500 rpm for 10 minutes before being redispersed in 20 mL of distilled water.

### **Conjugation of Bisphosphonate**

2mg/ml of Zolendronic acid was added to one part of the sample gold nanoparticle extract and left to stir overnight.

### **Antimicrobial Activity**

Biosynthesized nanoparticles are used in a wide range of biomedical applications. Membrane damage is one of the most common causes of nanoparticle antibacterial properties. Using the agar well diffusion method, the green synthesized bisphosphonate conjugated gold nanoparticles were tested against common oral pathogens such as *Candida albicans*, *Enterococcus faecalis*, *Staphylococcus aureus*, and *Lactobacillus*. The test organisms (*S. mutans* and

Lactobacillus) were grown in nutrient broth and kept on agar slants for the study. *Candida albicans* were grown on Rose Bengal agar, which is a yeast-specific medium. Using a sterile cotton swab, the freshly cultured strains were grown and uniformly spread over petri dishes containing MHS agar (Mueller Hinton 2 agar + 5% sheep blood). With the help of a steel borer, agar wells measuring 6.0 mm in diameter were punched into the culture plate containing the test microorganisms 35.

A micropipette was used to fill the agar wells with 20 $\mu$ L of different concentrations of nanoparticles (50,100,150g/ml). As a positive control, 20 $\mu$ L of standard antibiotics (Ampicillin) were used. The diameter of the inhibition zone was measured in millimeters after a 24-hour incubation period at 37°C (mm). All of the tests were performed three times.

### **Cytotoxicity**

The eggs of brine shrimp are purchased to perform the cytotoxic assay on brine shrimp. The eggs are then kept at a temperature of 28°C. Artificial seawater and a 37°C light source are used to hatch eggs. This method was tested in 15 well plates (Figure 1) The newly hatched Nauplii are selected and transferred to each well using a Pasteur pipette. The Gold nanoparticles with and without Bisphosphonate conjugation were introduced into each of the wells of varying concentrations of 5,10,15,25  $\mu$ L is added to each well, and the volume is adjusted. After 24 hours, the brine shrimp are removed from the 15 well plates and counted with a magnifying glass. After a 24-hour incubation period, the percentage of dead shrimp in each well is calculated. The number of motile nauplii was calculated to assess the cytotoxicity of the nanoparticles

Viability was calculated per well by

$$\text{Viability (in \%)} = \frac{\text{live brine shrimp after exposure}}{\text{live brine shrimp before exposure}} * 100\% \quad (46)$$

### **Result and Discussion**

#### ***Structural characterization of nanoparticles:***

#### **Visual colour change:**

The color change of the reaction mixtures from light yellow to yellow, dark-purple, and dark brown, respectively, could indicate the biosynthesis of Au nanoparticles in the current experiment. In the current experiment, the visual color change from yellow to dark purple was formed in 6 hours. The reduction confirmation of Au<sup>++</sup> to Au<sup>0</sup>, as indicated by the solution's color changing from light brown to dark brown. The brown color variation indicates an incomplete reduction of less concentration in the plant extract solution, whereas the formation of dark brown color at high plant extract concentrations revealed a complete reduction reaction.

#### **UV Spectroscopy Analysis:**

In the presence of incident photons, Au nanoparticles displayed the surface plasmon resonance (SPR) band as a result of the metal's conduction and free

band electrons collectively oscillating. The intensity of the SPR band is primarily determined by the nature of the nanoparticles used in the synthesis, as well as their composition. Furthermore, UV-vis spectroscopy is a key tool for determining the nature of synthesized Au. The analysis was carried out every one hour to determine the changes. The analysis showed a consistent peak after 1 hr of preparation at 540 nm was constantly observed after 2 hours of the preparation of the sample (Figure 1a and 1b).

### **TEM and crystallographic analysis**

The TEM images and EDX spectra of biosynthesized Au nanoparticles showed that the particles are narrow in size and spherical in shape with a diameter in the range of 10–50 nm (Figure 2). However, some froth was noticed on the surface of these obtained nanoparticles, which could be attributed to the different types of phytochemicals present in the plant extract. FTIR analysis confirmed the presence of a huge amount of phytochemicals in the plant extract which can prevent the nanoparticles from agglomeration and helps in the production of stable nanoparticles. There was no other defined morphological difference observed in the preparation of Au nanoparticles.

### **Antibacterial Activity of Biosynthesized Bisphosphonate Conjugated Gold Nanoparticle Using *Cissus Quadrangularis* Extract of *Staphylococcus Aureus*, *Lactobacillus*, and *Candida Albicans***

Antibacterial activity of biosynthesized bisphosphonate conjugated gold nanoparticle using *Cissus quadrangularis* extract on *Staphylococcus aureus*, *Lactobacillus*, and *Candida albicans* using agar well diffusion method was performed. This method is widely used to evaluate the antimicrobial activity of plants or microbial extracts. It is qualitative, easy to perform, and simple. The agar plate surface was inoculated by spreading a volume of the microbial inoculum over the entire agar surface. Then, a hole with a diameter of 6 mm punched aseptically with a sterile cork borer or a tip, and a volume of 20  $\mu$ L of the nanoparticle sample at desired concentration was introduced into the well. Then, agar plates were incubated under suitable conditions depending upon the test microorganisms. The nanoparticle sample diffuses in the agar medium and inhibits the growth of the microbial strain tested.

In this study, four different concentrations of the nanoparticles were studied (25, 50, 100, 150  $\mu$ g/ml). The diameter of the zone of inhibition increased with an increase in the concentration of the nanoparticles, against both *S. aureus*, *Lactobacillus*, and *Candida albicans*. Whereas, the diameter of the zone of inhibition against *Enterococcus fecalis* showed no change with the concentration of the nanoparticles. The zone of inhibition (in millimeters) of gold nanoparticles of varying concentrations, against *S. aureus*, *Lactobacillus*, and *Candida albicans* are represented in Table 1. Au nanoparticles have good antibacterial activity against *Streptococcus mutans* (150  $\mu$ g/ml – 26 mm zone of Inhibition), *Lactobacillus* (150  $\mu$ g/ml – 26 mm zone of Inhibition), and *Candida albicans* (150  $\mu$ g/ml – 10 mm zone of Inhibition).

## **Cytotoxicity assessment using brine shrimp lethality assay**

Cytotoxicity of the prepared nanoparticles was assessed using Brine Shrimp (*Artemia salina*) Lethality Assay. It has been demonstrated that the early developmental stages of *Artemia salina* are highly vulnerable to toxins. The lethality was found to be directly proportional to the concentration of the nanoparticles. Gold nanoparticles both with and without Bisphosphonate conjugation showed mortality of 10% at 20 and 25 ug/ml (Table 2).

## **Conclusion**

The present study revealed that green preparation of gold nanoparticles is achievable in a simple and eco-friendly manner using *Cissus quadrangularis* extract. The gold nanoparticles were assessed for their Antibacterial activity against *S.mutans*, *Lactobacillus*, and *C.albicans*. Cytotoxicity of the nanoparticles was assessed using Brine Shrimp Lethality Assay. Gold nanoparticles have good antibacterial activity against *Streptococcus mutans* (150 ug/ml – 17 mm zone of Inhibition) & *Lactobacillus* (150 ug/ml – 20 mm zone of Inhibition) and showed 20% lethality at 50ul concentration.

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**Conflict of interest:** There are no conflicts of interest.

## **Author contribution:**

Dr. Sruthi Harikrishnan: Methodology, Software, Validation, Formal Analysis, Investigation, Writing – Original Draft Preparation.

Dr.Navaneethan R : Conceptualization, Resources,Supervision, Project Administration, Writing – Review & Editing

Dr.Rajesh kumar: Conceptualization, Data Curation , Writing – Review & Editing.

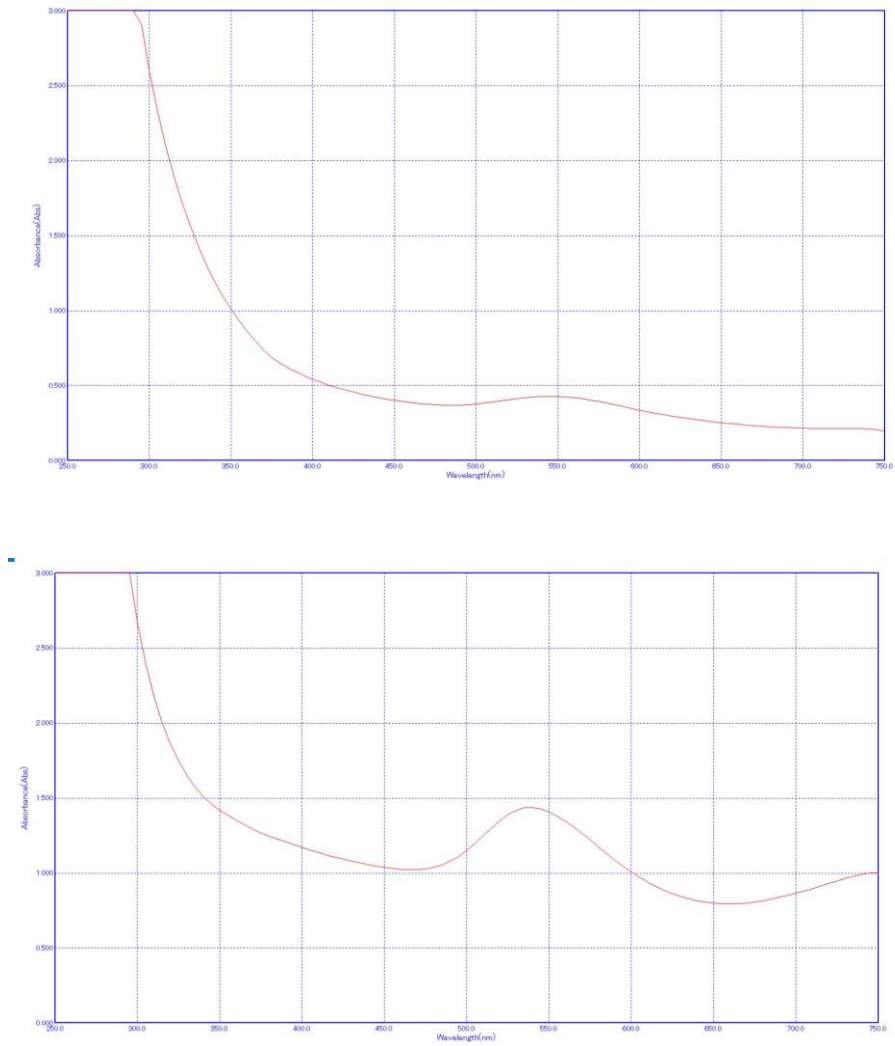
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Figures 1a and 1b: Shows the formation of the peak from 0 hr to 2 hr indicating the formation of the gold nanoparticles

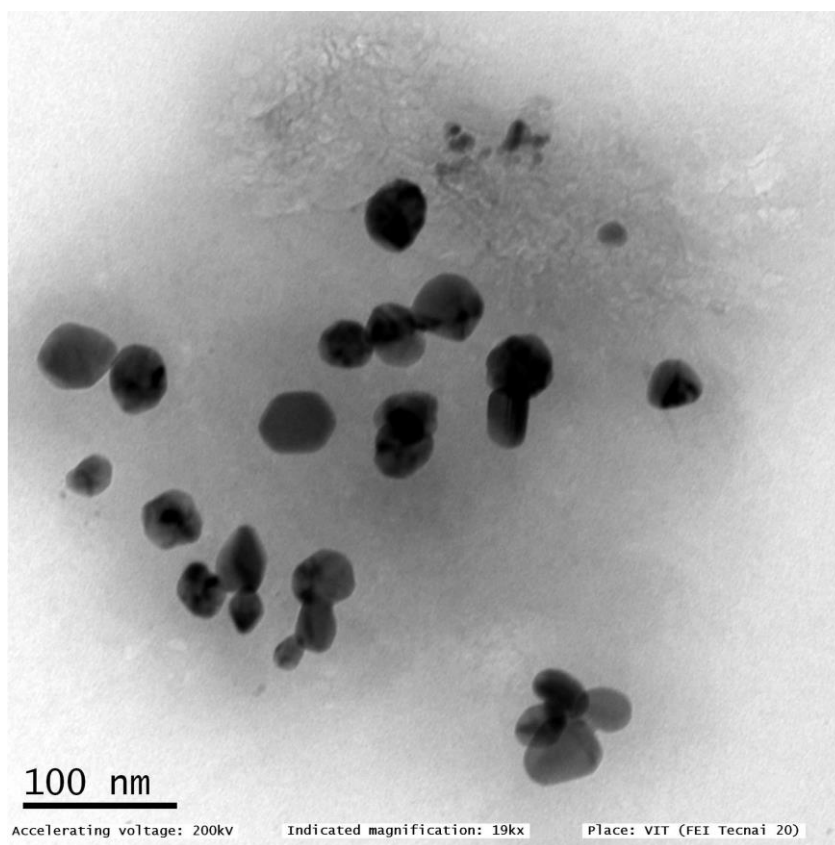


Figure 2: Transmission Electron Microscopic images of Gold nanoparticles. AuNPs synthesized in this study appear spherical, smooth, and measuring approximately 10–50 nm.

Table 1: Antimicrobial activity of Bisphosphonate conjugated gold nanoparticles at various concentrations

Nanoparticles	Organisms & zone of inhibition for varying concentrations of nanoparticles (ZOI) in millimeter (mm)														
	staphylococcus aureus					Lactobacillus					Candida albicans				
Bisphosphonate conjugated gold nanoparticles	25µg/ml	50µg/ml	100µg/ml	150µg/ml	control	25µg/ml	50µg/ml	100µg/ml	150µg/ml	Control	25µg/ml	50µg/ml	100µg/ml	150µg/ml	Control
	9	9	9	26	24	9	9	9	26	26	9	9	9	10	12

Table 2: Calculation of cytotoxicity at various concentrations of nanoparticles

CONCENTRATION(µg/mL)	No. of live Nauplii (Day 1)	No. of live Nauplii (Day 2)	% DEAD
CONTROL	10	10	0
GOLD nanoparticles WITHOUT BISPHOSPHONATE			
5 µg/ml	10	10	0
10 µg/ml	10	10	0
15 µg/ml	10	10	0
20 µg/ml	10	9	10%
25 µg/ml	10	9	10%
GOLD NANOPARTICLES WITH BISPHOSPHONATE			
5 µg/ml	10	10	0
10 µg/ml	10	10	0
15 µg/ml	10	10	0
20 µg/ml	10	8	20%
25 µg/ml	10	9	10%