

How to Cite:

AbdulAmeer, A. K., & Zaid, N. W. (2022). General and Progressive Semen Motility Affected by using Black Currant (*Prunus cerasus*) Extracts in Rams . *International Journal of Health Sciences*, 6(S7), 3386–3394. <https://doi.org/10.53730/ijhs.v6nS7.12477>

General and progressive semen motility affected by using black currant (*Prunus Cerasus*) extracts in rams

Ahzan Kh. AbdulAmeer

Faculty of Veterinary Medicine, University of Kufa, Iraq

Email: ahzank.alfatlawe@uokufa.edu.iq

Nazih Wayes Zaid

Department of Surgery and Obstetrics, College of Veterinary Medicine, University of Baghdad, Baghdad, Iraq

E.mail: nazihwayeszaid@covm.uobaghdad.edu.iq

Abstract--This study was designed in order to investigate and determine the effect of black current (*Prunus cerasus*) extract on semen general and progressive motility of the rams. Twelve mature healthy rams was used in this study aged between 2-3 years, kept in the animal farm of the College of Veterinary Medicine, Al-Kufa University. Semen ejaculates was collected using electroejaculator methods once weekly a total of 36 ejaculates were collected among the period of the study. The black current (*Prunus cerasus*) extract was prepared by using maceration and added to the semen in five doses groups (0.25 gm/ml as G1, 0.5 gm/ml as G2, 1 gm/ml as G3, 2 gm/ml as G4, 4 gm/ml as G5) and the positive control group left without any addition. The treatment samples were evaluated under cooling for five consecutive days, the general and progressive motility were analysis using Computer Assisted Sperm Analysis (CASA). The results of the study clearly indicated that there were a significant differences ($P<0.05$) in G1 and G2 than other groups in five days of examination, with the preference to the G1 group. We conclude that the 0.25 and 0.5 gm/ml of black current extract (*Prunus cerasus*) should be used in cooling process as a part of artificial insemination in sheep.

Keywords---semen, general motility, progressive motility, black current, rams.

Introduction

Antioxidants are the compound that prevent the creation of ROS or oppose their actions (Bansal and Bilaspuri, 2008). Previous studies have shown that inclusion of antioxidants in cryopreservation extenders increases factors such as motility and membrane integrity after thawing in human, ram, goat, bull, canine and boar sperm (Bucak *et al.*, 2010). Supplementation with antioxidants prior to the cryopreservation process may improve cryopreservation methods used in the sheep industry (Bucak *et al.*, 2010). The limited ability of sperm to biosynthesize, coupled with the reduction in antioxidant concentrations present in the semen after dilution, reduces the constructive effect of these endogenous anti-oxidative defenses (Allai *et al.*, 2018). Therefore, even a small addition of antioxidants to cryopreservation media may have a positive effect on sperm function (Allai *et al.*, 2018). Natural antioxidants are compounds that are obtained from natural sources such as herbs, seeds, vegetables, spices and fruits (Velasco and Williams, 2011). These natural antioxidants are often phenolic compounds or vitamins that have an appropriate molecular structure allowing them to chemically scavenge free radicals (Liang *et al.*, 2012). In plants, phenolic compounds have numerous biological roles such as antioxidants, anti-bacterial and anti-inflammatory (Manthey and Grohmann, 2001). Antioxidants obtained from natural sources such as green tea and rosemary have the ability to enhance sperm integrity, which minimizes the damage that occurs during storage. There are limited studies on the effect of medical plants extract as antioxidant on spermatozoa quality during and after cooling. One of well-known medicinal plants is black currant (*Prunus cerasus*), it has an excellent antioxidant activity (Piccolella *et al.*, 2008; Kirakosyan *et al.*, 2010; Damar *et al.*, 2012 and Xu *et al.*, 2016) respectively, anti-inflammatory activity (Desjardins *et al.*, 2012) as well as anti-carcinogenic activity (Al Alawi *et al.*, 2020). This study was design in order to investigate the effect of black currant (*Prunus cerasus*) adding to ram semen on general and progressive motility.

Materials and methods:

The extraction of plant was done in the artificial insemination and theriogenology laboratory, inside Veterinary Medicine College, University of Kufa, Iraq. The method of choice for the aqueous extraction was done by using maceration of the crude extracts, due to its beneficial in extracting of the thermolabile and thermostable components (Al Alawi *et al.*, 2020). The edible part of the black current Fruits were separated manually from the seeds and chopped into small pieces. The black current plant 200 gram was treated with 300 ml of water and mixed very well at room temperature. The content of bottle then was filtered through a Buchner filtration system and the water was removed by exposure to 45 °C to evaporate the water (Bustani, 2020). The remaining plant was treated again with water and the process was repeated for twice times. The extract was collected and stored in room temperature for later use.

The extender was divided into six aliquots dose of aqueous plant extract as 0% (C), 0.25 mg/ml (G1), 0.5 mg/ml (G2), 1 mg/ml (G3), 2 mg/ml (G4) and 4 mg/ml (G5), the solution had normal pH ranged between 6.8-7.2 and 425mOsm osmolality. The general and progressive motility of the semen was evaluated at

day 1, 2, 3, 4, and 5. The extender was stored after preparation at -20 °C. The extender was used in this study is a Tris- based extender includes: (for 100 ml) LDL 8%, tris: 2.40 g, fructose: 1 g, citric acid: 1.50 g, gentamycin 25 mg, penicillin 50000 IU, and D. W. until 100 ml (Baiee *et al.*, 2017). The extract was added to the extender with different doses as mentioned before and one group kept without adding the extract as a control.

Samples of semen were collected from 12 mature ram aged between 2–3 years using electro-ejaculator method (mini tube; Germany) every two weeks. Thirty six ejaculates of fresh semen were collected and evaluated at 36 °C, general and progressive motility of sperm was done using **Computer Assisted Sperm Analysis (CASA)**. Selected samples should be $\geq 70\%$ sperm motility and $\geq 80\%$ normal morphology in order to be used in this study.

The extended semen groups were stored in a refrigerator at 5 °C for five days. Sperm general and progressive motility were evaluated at 1, 2, 3, 4, and 5 days of chilling. The final concentration of semen in the chilled experiment was approximately 80×10^6 sperm/ml.

The statistical analysis was done using ANOVA and LSD was used to determine the significant differences among means at level of $P < 0.05$.

Results

According to Figure (1), the outcomes of the result that illustrated the value of general sperm motility in the different concentration of aqueous plant extract of *Prunus cerasus* (C, G1, G2, G3, G4 and G5). Moreover, the result showed superiority of G1 (0.25 mg/ml) among all other groups (Figure 1 and 2). In additionally the outcome showed significant increase ($P < 0.05$) in the general motility of G1 at 3rd, 4th and 5th day of preservation that compared with control and other groups (Figure 1 and 2). In other hand, the general and progressive motility of G2 (0.5 mg/ml) group was higher than control in 3rd, 4th and 5th of day of preservation (Figure 1 and 2). Generally, the general and progressive motility of both G1 and G2 group higher significantly ($P < 0.05$) than G4 and G5 groups in all day of preservation (Figure 1 and 2). The motility of G1 was increased significantly ($P < 0.05$) than G3 while that non-significant different between G2 and G3 groups (Figure 1 and 2).

Row stats of Generally Motility

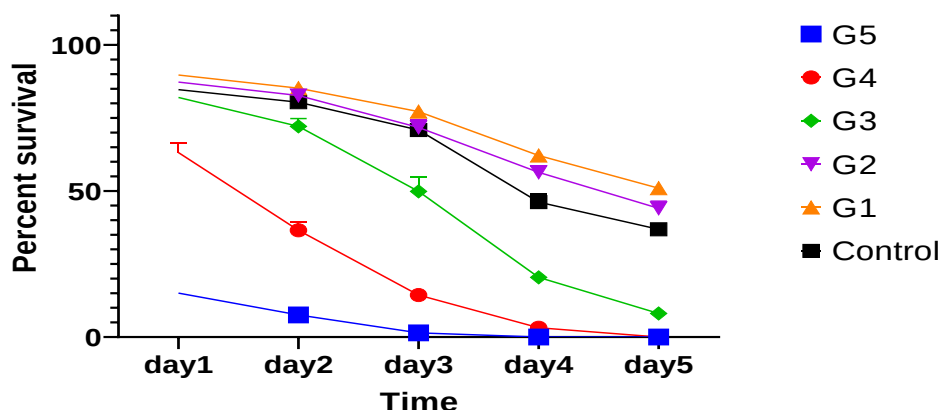


Figure 1: Comparison among groups of sperm general motility % during 5 day survival.

Progressive Motility

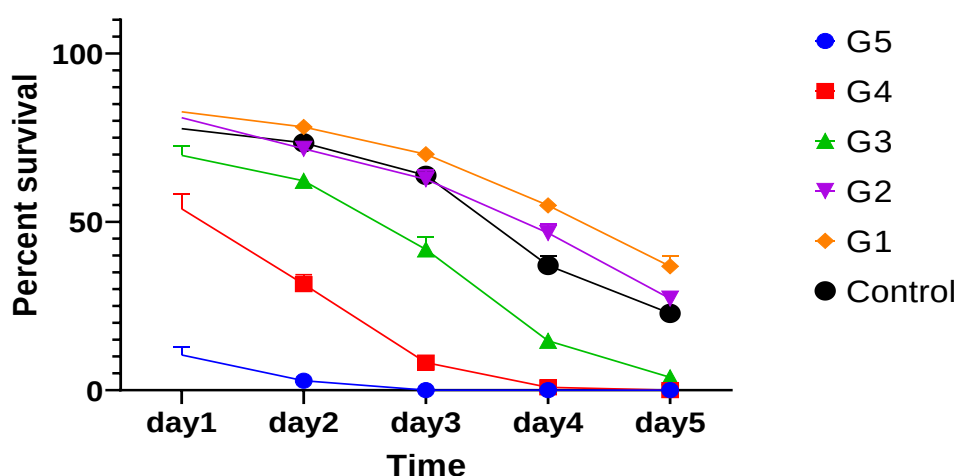


Figure 2: Comparison among groups of sperm progressive motility during 5 day survival.

Discussion

The current study found that semen diluents with 0.25 mg/ml aqueous extract of *Prunus cerasus* significantly improved the cooled sperm parameters and it was in line with the gold standard diluents which are tris-egg yolk diluents. Thus, addition of 0.25 mg/ml of aqueous extract enhanced the functional parameters and in vivo fertility. On the other hand, oxidative stress, lipid peroxidation, and thawing effect are main causes of spermatozoal damage through preservation (Barman *et al.*, 2011 and Rizkallah *et al.*, 2022) as approbate our result in control group. Furthermore, the reduced of temperature during chilling processing which

preserved at 5 °C or less inflicts sub-lethal damage to spermatozoa, compromising sperm quality and the success of artificial breeding as described by previous studies (Mohamed *et al.*, 2020 and Bustani and Baiee 2021). The earlier studies have concluded effect of chilling temperature on the sperm motility in both general and progressive motile of rat (Varisli *et al.*, 2009), bull (Januskauskas *et al.*, 1999), rabbit (Rosato *et al.*, 2011), human (Verheyen *et al.*, 1993) and, ram (Kulaksiz *et al.*, 2012) which illustrated the temperature related in the organization of structural components of the plasma membrane have profound effects upon its properties, associated with changes in permeability, changes in membrane fluidity (Ortega *et al.*, 2019), dis-regulation of intracellular Ca^{+} influx (Yeste *et al.*, 2018) and changes in enzyme activity (Palhares *et al.*, 2021) finally on the ability of sperm capacitation. From the Figure (1 and 2) the result showed the improved effect of *Prunus cerasus* (G1) on the sperm motility (general and progressive). Which the past studies proved the antioxidant effect of *Prunus cerasus* on different tissue and cells (Sokół-Łętowska *et al.*, 2020), furthermore, the contain of high amount of sugars (glucose and fructose) since the sugar (glucose or fructose) offers a good protection during preservation of semen and this might be due to be a source of energy (Williams and Ford, 2001). Sperm motility doesn't depend on mitochondrial to provide the energy that used to motility action. With this in mind, studies suggested that glycolysis is an alternative pathway to produce ATP in the cytoplasm of sperm cell (Nascimento *et al.*, 2008); since, it was reported that glycolytic enzymes and an ADP/ATP carrier protein are found in the fibrous sheath of sperm tail (Krisfalusi *et al.*, 2006 and Kim *et al.*, 2007). Therefore, spermatozoa could use glycolysis to produce ATP that can be used for motility. A handful of studies reported that sperm motility was significantly increased when incubated in diluents containing sugar like glucose (du Plessis *et al.*, 2015). Generally, glycolysis and oxidative phosphorylation provide energy for sperm motility (Qiu *et al.*, 2016). As well as, sugars prevent cooling damage of sperm by affecting the pattern of ice-crystal formation and the width of the channels of unfrozen extender (Nicolajsen and Hvidt, 1994) and reduce the salt concentration of the extender that can reduce the injury due to the extender solution effects. Recent studies were indicated that spermatozoa motility depends greatly on the energy source in an extender in the form of adenosine triphosphate (ATP), which is produced from metabolism, and therefore requires a constant supply for cell function and survival (Dorado *et al.*, 2019). This study result finding broadly supports the work of other studies in this area linking motility with source of energy and production of ATP which our result showed increased significantly in the general and progressive motility in addition of aqueous semen extender as showed in Figure (1 and 2). Moreover, the figures illustrated the superiority of G1 significantly among all groups, This study supports evidence from previous observations (Alenezy *et al.*, 2019; Asadzadeh *et al.*, 2021 and El-Harairy *et al.*, 2018) uses of natural source of material in the extender of semen for chilling and cryopreservation of the bull sperm, in the same area the current study found that used of vitamins are promote and increased sperm parameter (general and progressive motility) is support our result (Espina *et al.*, 2021). Also the season in Iraq had a significant effect upon animal testis specially the summer season which show the lowest reproductive activity (Ibrahim and Zaid, 2017).

It can be notable that works precisely at semen diluents with 0.25 mg/ml aqueous extract of *Prunus cerasus* (G1) and based on the present data it provides a good opportunity to improve sperm quality. However, high concentration at 2 mg /ml and 4 mg /ml of *Prunus cerasus* G4 or G5 did not improve post-thawed semen quality. This was might due to the effect of disaccharide that found in small amount in the extract of dates. Since, a study on epididymal spermatozoa of red deer found that mono-saccharides had positive effect than di-saccharides or the tri-saccharide (raffinose) when supplementing diluents for cryopreservation (Fernández-Santos *et al.*, 2007). In addition, the negative effect of higher concentration of AED or SED at dose of 2 g / 100 ml was concomitant with previous studies where high concentration of disaccharide have been used to limit their dosages effect on post-thawed sperm quality (Molinia *et al.*, 1994).

Conclusion

From the previous results we conclude that the black currant (*Prunus cerasus*) extracts had a remarkable effect on semen general and progressive motility of the rams on 0.25 and 0.5 gm/ml during cooling with good longevity effect, and it should be used in artificial insemination programs of sheep.

Ethics approval and consent to participate: Not applicable.

Availability of data and material: The data that support the findings of this study are available from the corresponding author, upon reasonable request.

Competing interests: None of the authors have any conflict of interest to declare.

Funding Statement: This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Authors' contributions: All authors contributed equally.

ORCID link of the corresponding author: [https://orcid.org/ 0000-0002-6996-9368](https://orcid.org/0000-0002-6996-9368)

References:

- Bansal K, and Bilaspuri GS, Effect of ferrous sulphate and ascorbic acid on motility, viability and lipid peroxidation of crossbred cattle bull spermatozoa. *Animal*. 2008; 2(1): 100-4.
- Bucak MN, Sariozkan S, and Tuncer PB, The effect of antioxidants on post-thawed Angora goat (*Capra hircus ancryrensis*) sperm parameters, lipid peroxidation and antioxidant activities. *Small Ruminant Res.* 2010; 89(1): 24–30.
- Allai L, Benmoula A, da Silva MM, Nasser B, and Al Amiri B, Supplementation of ram semen extender to improve seminal quality and fertility rate. *Animal Reproduction Science*. 2018; 192: 6-17.
- Velasco V, and Williams P, Improving meat quality through natural antioxidants. *Chilean J Agri Res.* 2011; 71(2): 313-322.
- Liang J, Li JK, Zhao W, and Liu YL, Effect of pomegranate peel polyphenols on lipid peroxidation in vitro. *J Food Sci Biotech.* 2012; 31: 159-165.

- Manthey J, and Grohmann K, Phenols in Citrus Peel Byproducts. Concentrations of Hydroxycinnamates and Polymethoxylated Flavones in Citrus Peel Molasses. *J Agri Food Chem.* 2001; 49(7): 3268-3273.
- Piccolella S, Fiorentino A, Pacifico S, D'Abrosca B, Uzzo P, and Monaco P, Antioxidant properties of sour cherries (*prunus cerasus* L.): Role of color less phytochemicals from the methanolic extract of ripe fruits. *J Agric Food Chem.* 2008; 56(6): 1928–1935.
- Kirakosyan A, Seymour EM, Noon KR, Llanes DEU, Kaufman PB, and Warber SL, Interactions of antioxidants isolated from tart cherry (*Prunus cerasus*) fruits. *Food Chem.* 2010; 122 (1): 78–83.
- Damar I, and Eksi A, Antioxidant capacity and anthocyanin profile of sour cherry (*prunus cerasus* L.) juice. *Food Chem.* 2012; 135 (4): 2910–14.
- Xu Y, Cai F, Yu Z, Zhang L, Li X, Yang Y, and Liu G, Optimisation of pressurised water extraction of polysaccharides from blackcurrant and its antioxidant activity. *Food chem.* 2016; 194: 650-8.
- Desjardins J, Tanabe S, Bergeron C, Gafner S, and Grenier D, Anthocyanin-rich black currant extract and cyanidin-3-O-glucoside have cytoprotective and anti-inflammatory properties. *J med food.* 2012; 15(12): 1045-1050.
- Al Alawi R, Alhamdani MSS, Hoheisel JD, and Baqi Y, Antifibrotic and tumor microenvironment modulating effect of date palm fruit (*Phoenix dactylifera* L.) extracts in pancreatic cancer. *Biomed and Pharm.* 2020; 121: 109522.
- Bustani GSAA, Effect of *Phoenix dactylifera* extracts supplemented to tris on preserved sperm parameters and pregnancy rates in bull. Thesis of Master Science in Physiology- College of Veterinary Medicine- University of Kufa. 2020.
- Baiee FH, Wahid H, Rosnina Y, Ariff OM, Yimer N, Salman H, and Khumran AM, Hypo-Osmotic Swelling Test Modification to Enhance Cell Membrane Integrity Evaluation in Cryopreserved Bull Semen. *Pertanika J Trop Agri Sci.* 2017; 40(2): 257-268.
- Barman K, Asrey R, and Pal RK, Putrescine and carnauba wax pretreatments alleviate chilling injury, enhance shelf life and preserve pomegranate fruit quality during cold storage. *Sci Hort.* 2011; 130(4): 795-800.
- Rizkallah N, Chambers CG, de Graaf SP, and Rickard JP, Factors Affecting the Survival of Ram Spermatozoa during Liquid Storage and Options for Improvement. *Animals.* 2022; 12(3): 244.
- Mohamed MY, Mahdy TM, and Khalifa EI, The possibility of using citric phosphate dextrose in chilling ram semen instead of egg yolk and soybean lecithin to improve fertility. *J Ani Physio Ani Nutri.* 2020; 104(6): 1620-27.
- Bustani GS, and Baiee FH, Semen extenders: An evaluative overview of preservative mechanisms of semen and semen extenders. *Vet World.* 2020; 14(5): 1220.
- Varisli O, Uguz C, Agca C, and Agca Y, Effect of chilling on the motility and acrosomal integrity of rat sperm in the presence of various extenders. *J Amer Associ Lab Ani Sci.* 2007; 48(5): 499-505.
- Januskauskas A, Gil J, Söderquist L, Hrd MGM, Hrd MC, Johannisson A, and Rodriguez-Martinez H, Effect of cooling rates on post-thaw sperm motility, membrane integrity, capacitation status and fertility of dairy bull semen used for artificial insemination in Sweden. *Theri.* 1999; 52(4): 641-658.

- Rosato MP, and Iaffaldano N, Effect of chilling temperature on the long-term survival of rabbit spermatozoa held either in a tris-based or a jellified extender. *Reprod domes ani*. 2001; 46(2): 301-8.
- Verheyen G, Pletincx I, and Van Steirteghem A, Andrology: Effect of freezing method, thawing temperature and post-thaw dilution/washing on motility (CASA) and morphology characteristics of high-quality human sperm. *Human Reprod*. 1993; 8(10): 1678-84.
- Kulaksiz R, Çebi Ç, and Akcay E, The effect of different extenders on the motility and morphology of ram sperm frozen or stored at 4 °C. *Turkish J Vet Ani Sci*. 2012; 36(2): 177-182.
- Ortega-Morales LD, Alcantar-Rodriguez A, Espejel MC, and Medrano A, The effect of non-traditional cooling on dog sperm cryosurvival and ability to perform the acrosome reaction. *Aust J Vet Sci*. 2019; 51(2): 73-82.
- Yeste M, Castillo-Martín M, Bonet S, and Rodríguez-Gil JE, Impact of light irradiation on preservation and function of mammalian spermatozoa. *Ani Repro Sci*. 2018; 194: 19-32.
- Palhares PC, Assis IDL, Machado GJ, de Freitas RM, de Freitas MB, Paula DA, Carneiro WF, Motto NC, and Murgas LD, Sperm characteristics, peroxidation lipid and antioxidant enzyme activity changes in milt of *Bryconorbignyanus* cryopreserved with melatonin in different freezing curves. *Therio*. 2021; 176: 18-25.
- Sokół-Łętowska A, Kucharska AZ, Hodun G, and Gołba M, Chemical Composition of 21 Cultivars of Sour Cherry (*Prunus cerasus*) Fruit Cultivated in Poland. *Molecules*. 2020; 25(19): 4587.
- Williams AC, and Ford WCL, The role of glucose in supporting motility and capacitation in human spermatozoa. *J Andro*. 2001; 22(4): 680-95.
- Nascimento JM, Shi LZ, Tam J, Chandsawangbhuwana C, Durrant B, Botvinick EL, and Berns MW, Comparison of glycolysis and oxidative phosphorylation as energy sources for mammalian sperm motility, using the combination of fluorescence imaging, laser tweezers, and real-time automated tracking and trapping. *J Cell Physio*. 2008; 217(3): 745-51.
- Krisfalusi M, Miki K, Magyar PL, and O'Brien DA, Multiple glycolytic enzymes are tightly bound to the fibrous sheath of mouse spermatozoa. *Biol Reprod*. 2006; 75(2): 270-8.
- Kim YH, Haidl G, Schaefer M, Egner U, Mandal A, and Herr JC, Compartmentalization of a unique ADP/ATP carrier protein SFEC (Sperm Flagellar Energy Carrier, AAC4) with glycolytic enzymes in the fibrous sheath of the human sperm flagellar principal piece. *Develo Biol*. 2007; 302(2): 463-76.
- du Plessis SS, Agarwal A, Mohanty G, and Van der Linde M, Oxidative phosphorylation versus glycolysis: what fuel do spermatozoa use?. *Asian J Andro*. 2015; 17(2): 230- 5.
- Qiu JH, Li YW, Xie HL, Li Q, Dong HB, Sun MJ, Gao WQ, and Tan JH, Effects of glucose metabolism pathways on sperm motility and oxidative status during long-term liquid storage of goat semen. *Therio*. 2016; 86(3): 839-49.
- Nicolajsen H, And Hvidt A, Phase behavior of the system trehalose-NaCl-water. *Cryobiology*. 1994; 31(2): 199-205.
- Dorado J, Hidalgo M, Acha D, Ortiz I, Bottrel M, Azcona F, Carrasco JJ, Gomez-Arrones V, and Demyda-Peyrás S, Cryopreservation of Andalusian donkey (*Equus asinus*) spermatozoa: Use of alternative energy sources in the freezing

- extender affects post-thaw sperm motility patterns but not DNA stability. *Ani Reprod Sci.* 2019; 208: 106-26.
- Alenezzy ES, Barakat IA, and Al Musayeib NM, Effect of wild marjoram (*Origanum vulgare*) plant extracts on capacitation of sheep spermatozoa in vitro. *Advan Bio Biotech.* 2019; 10(4): 82-97.
- Suryasa, I. W., Rodríguez-Gámez, M., & Koldoris, T. (2022). Post-pandemic health and its sustainability: Educational situation. *International Journal of Health Sciences*, 6(1), i-v. <https://doi.org/10.53730/ijhs.v6n1.5949>
- Asadzadeh N, Abdollahi Z, Esmaeilkhanian S, and Masoudi R, Fertility and flow cytometry evaluations of ram frozen semen in plant-based extender supplemented with Mito-TEMPO. *Anim Reprod Sci.* 2021; 233: 106836.
- El-Harairy MA, Khalil WA, Khalifa EI, and Saber AA, Effect of Propolis Ethanolic Extract Supplementation to Ram Semen Extenders on Sperm Characteristics, Lipid Peroxidation and some Enzymatic Activities in Seminal Plasma in Chilled Semen. *J Anim Poul Prod.* 2018; 9(4): 235-43.
- Espina-Ávila JA, Magaña-Monforte JG, Aké-Villanueva JR, and Aké-López JR, Effect of the use of vitamin “E” in the diluent on the viability of ram sperm. *Trop Anim Heal Prod.* 2021; 53(2): 1-6.
- Ibrahim NS, and Zaid NW, Testosterone role during seasons changes in the dogs testes. *The Iraqi Journal of Veterinary Medicine.* 2017; 41(1): 125-30.
- Fernández-Santos MR, Estesó MC, Montoro V, Soler AJ, and Garde JJ, Cryopreservation of Iberian red deer (*Cervus elaphus hispanicus*) epididymal spermatozoa: effects of egg yolk, glycerol and cooling rate. *Therio.* 2006; 66(8): 1931-42.
- Molinia FC, Evans G, and Maxwell WMC, In vitro evaluation of zwitterion buffers in diluents for freezing ram spermatozoa. *Reprod Nutri Develop.* 1994; 34(5): 491-500.