

How to Cite:

Aboobecker Siddique, P. A., Gaur, A., Bajpai, A., John, A., & John, A. (2022). Evaluation of hepatoprotective potential of *rubia cordifolia* in experimentally (carbon tetrachloride) induced hepatotoxicity in albino rats. *International Journal of Health Sciences*, 6(S4), 804–814. <https://doi.org/10.53730/ijhs.v6nS4.5708>

Evaluation of hepatoprotective potential of *rubia cordifolia* in experimentally (carbon tetrachloride) induced hepatotoxicity in albino rats

Mr. Aboobecker Siddique P. A

Assistant Professor, Department of Pharmacology

Dr. Anand Gaur

Assistant Professor, Department of Pharmacology, N. C Medical College & Hospital, Israna, Panipat, Haryana

Dr Anurag Bajpai

Professor and HOD, Department of Pharmacology, N. C Medical College & Hospital, Israna, Panipat, Haryana

Dr. Abhay John

Associate Professor, Department of Pharmacology, Amaltas Institute of Medical Science, Dewas

Abstract--Liver is one of the largest glands of the body and the main site for intense metabolism and excretion. Liver disease is a term for a collection of conditions that adversely affect the cells, tissues, structures, or functions of the liver. Treatment of liver diseases with various drugs develops risk of toxicity. *Rubiocordifolia* is an herbal plant has, been claimed to have hepatoprotective potentials. The present study was conducted to explore the hepatoprotective activity of aqueous extract of *Rubiocordifolia* against carbon tetrachloride induced hepatotoxicity in albino rats. This experimental study was conducted after getting approval from institutional animal ethics committee using, routes of *Rubiocordifolia* in wistar albino rats (150-200g) of either sex. The hepatoprotective activity was evaluated by using histo-pathological examination. Animals were divided in to five groups with six animals in each group. Group-1 was given Normal saline (1ml/kg/day), Group-2 was injected with carbon tetrachloride (1ml/kg) i.p. only once to produce hepatotoxicity. Group-3 and Group-4 were given *Rubiocordifolia* (100mg/kg and 200mg/kg) orally every morning for 21 days followed by carbon tetrachloride (1ml/kg) i.p on 21st day (respectively). Group-5 was given standard drug Liv-

International Journal of Health Sciences ISSN 2550-6978 E-ISSN 2550-696X © 2022.

Corresponding author: Dr. Abhay John

Manuscript submitted: 27 Jan 2022, Manuscript revised: 18 Feb 2022, Accepted for publication: 9 March 2022

52(1mg/kg) i.p. for 21 days followed by carbon tetrachloride (1ml/kg) i.p on 21st day. Aqueous extract of roots of *Rubiacordifolia* in oral doses showed a significant effect on Histopathology. Normal hepatic lobule architecture is seen. Hepatocytes and their nuclei are well visible. Only cellular debris is seen and no hepatocytes with nuclei are discernible. It can be concluded that the Aqueous extract of roots of *Rubiacordifolia* is a potent hepatoprotective agent.

Keywords--*rubiacordifolia*, carbon tetrachloride, hepatoprotective, liv-52.

Introduction

Liver plays an important role in our body. To have a proper understanding of liver ailments, it is important to have a detailed knowledge of hepatic structure and function. [1] Liver disease is a term for a collection of conditions that adversely affect the cells, tissues, structures, or functions of the liver. Symptoms of liver disease may be acute, occurring suddenly, or chronic, developing slowly over a long period of time. Chronic liver disease is much more common than acute. [2] The rates of chronic liver disease for men are two times higher than for women. Liver disease may range from mild to severe depending on the type of disease present. [3]

CCI4 is the classic hepatotoxin, classified as an agent producing direct injury to the liver. Single dose lead promptly to zone 3 necrosis and steatosis, Prolonged administration can lead to cirrhosis and it also causes hepatic carcinoma in rats and mice. CCl₄ produces hepatic necrosis and steatosis in a variety of laboratory animals and in humans. [4] The hepatic damage is usually accompanied by lesions of other organs especially kidneys. Necrosis in zone 3 occurs in all species of mammals after administration of the agent by any route except direct injection into the portal venous system; the latter leads to massive necrosis. Hydropic degeneration (i.e. ballooning) and acidophilic degeneration in zone 3 precede the necrosis. These disappear when necrosis is maximal. [5]

The plant is commonly known as 'Indian Madder' and sold under the trade name 'manjistha'. It is a member of Rubiaceae family, distributed in hilly tracts of India up to 3750 m. It is a perennial, prickly, climbing herb with red rhizomatous base and roots. [6] Leaves variable, arranged four in a whorl, base slightly cordate, petioles are quadrangular, sometimes prickly on the angles, glabrous and shining. Stems is slender, rough, four angled with sharp recurved prickles on the ridges, which are often many yards long, becoming slightly woody at the base. Flowers are in cymes, greenish white. Fruits are didymous or globose, smooth, shining and purplish black when ripe. [7]

Aim

Evaluation of hepatoprotective potential of aqueous extract of *Rubia cordifolia* against carbon tetrachloride induced hepatotoxicity in albino rats.

Materials and Methods

This was an experiment study conducted on wistar rats in Department of Pharmacology, Index Medical College over a period of 2 year.

Healthy Albino wistar rats of either gender, weighing 150- 200g were obtained from CPCSEA approved Central Animal House of Medical College. The selected rats were housed in polpropylene cages under controlled conditions of temperature (25 °C) and alternating periods of light and darkness of 12 hours each. The rats had free access to standard rat pellet diet and tap water *ad libitum*. After one week of acclimatization, the animals were rendered suitable for study. Pregnant female rats were not included in the study.

Test compound

1. To induce hepatotoxicity, commercially available preparation of injectable Carbon tetrachloride was used.
2. Aqueous extract of *Rubia cordifolia*, to evaluate hepatoprotective activity.

Rubia cordifolia

Roots of *Rubia cordifolia* were dried under shade and ground to make a fine powder in a herb grinder. One gram of finely powdered plant material was extracted with 50 ml of water. The solution was filtered using Whatman filter paper no 1. Solution was evaporated under vacuum in a rotary evaporator to make the extract dry. This dried extract was stored at 0- 4°C and dissolved in water to make a solution of 1% concentration when ever required.

The animals were randomly divided into seven groups of six animals each. The groups are described as follows:

- Group- I: control group was administered 0.9% NaCl solution in a single oral dose of 1ml/kg body wt for 21 days.
- Group -II: In addition to pellet diet and tap water ad libitum, this group was injected with toxin CCl₄ (1ml/kg) i.p only once to produce hepatotoxicity on 21st day.
- Group-III: This group was given *Rubia cordifolia* orally (100 mg/kg) (Gilani AH et al.,1995). as a single dose per orally every morning for 21days followed by an injection of CCl₄ (1 ml/kg i.p.) on 21st day.
- Group -IV: This group was received *Rubia cordifolia* orally (200 mg/kg) as a single dose per orally every morning for 21days followed by an injection of CCl₄ (1 ml/kg i.p.) on 21st day.
- Group-V: This group received Liv.52 (1ml/kg) orally for 21 days followed by an injection CCl₄ (1 ml/kg i.p.) on 21st day.

Liv.52 and the test compound (*Rubia cordifolia*) were administered by gavage method with animals fasted 3-4 hours prior to and 1 hour after administration of test drugs to ensure proper absorption. After administration of carbon tetrachloride, animals of all the groups were fasted for 24 hours although which time water remained freely available during this period, Thereafter animals were sacrificed under Ketamine (75 mg/kg) and Diazepam (10 mg/kg) anaesthesia given intraperitoneally.

The liver was excised from the animals and washed with the normal saline. About one cm piece was cut and fixed in 10% neutral formalin for 12-24 hours. It was then dehydrated and cleared with ethanol and xylene respectively followed by embedding in paraffin wax from which blocks were prepared. Sections of 5m thickness were taken from the blocks using a microtome . These were processed in alcohol-xylene series and were stained with Harris haematoxylin and eosin stain [and subjected to histopathological examination.

Statistical Analysis

The data thus obtained was appropriately organized and analyzed by suitable statistical methods. i.e ; ANOVA.

Results

Aqueous extract of *Rubia cordifolia* showed effect on biochemical parameters.

Table 1 : Effect of Liv-52, *Rubia cordiafolia* in their respective doses on carbon tetrachloride induced changes in serum Alanine Transaminase (n=6).

TREATMENT (mg/kg)	Alanine transaminase(IU/L)(mean±SE)
Normal saline (1ml)	29.5±3.35
CCl ₄ (1)	433.5±48.67 [^]
LIV.52 (1)	140.7±8.1 [^]
<i>Rubia cordifolia</i> (100)	281±18.33 [*]
<i>Rubia codifolia</i> (200)	177.5±12.73 [*]

[‡]P< 0.05 as compare to ccl₄ treated group.

^{*}p< 0.01 as compare to ccl₄ treated group.

[^]P< 0.001 as compare to normal saline treated group.

ALT level in normal saline treated group was 29.5±3.35 IU/L. It was found to be significantly increased (p<0.001) with administration of CCl₄ to 433.5±48.67 IU/L. Pretreatment with known hepatoprotective preparation Liv.52 significantly (p<0.001) limited the rise in ALT levels after CCl₄ administration to 140.7±8.1 IU/L. With *Rubia cordifolia* there is dose depended limitation of ALT rise after CCl₄ administration. Although the dose of 100mg/kg for 21 days showed a significant limitation (p<0.01) of ALT rise (281±18.33) when compared to CCl₄ treated group but it did not match the efficacy of Liv. 52 treated group. However , in dose of 200 mg/kg for 21 days the *Rubia cordifolia* extract had much efficacy, in limiting the ALT rise after CCl₄ administration, to 177.5±12.73 IU/L, which was significant (p<0.01).

Table 2 : Effect of Liv-5, *Rubia cordifolia* in their respective doses on carbon tetrachloride induced changes in Alkaline Phosphatase (n=6).

TREATMENT (mg/kg)	Alkaline phosphatase (IU/L)(mean±SE)
Normal saline (1ml)	73.9±4.63
CCl ₄ (1)	600.42±52.9 [^]
LIV.52 (1)	198.9±11.6 [^]
<i>Rubia cordifolia</i> (100)	401±45.26 [£]
<i>Rubia codifolia</i> (200)	245.1±21.25 [*]

£P< 0.05 as compare to ccl₄ treated group.

*p< 0.01 as compare to ccl₄ treated group.

[^]P< 0.001 as compare to normal saline treated group.

ALP level in normal saline treated group was 73.9±4.63 IU/L. It was found to be significantly increased (p<0.001) with administration of CCl₄ to 600.42±52.9 IU/L. Pretreatment with known hepatoprotective preparation Liv.52 significantly (p<0.001) limited the rise in ALP levels levels after CCl₄ administration to 198.9±11.6 IU/L. With *Rubia cordifolia* there is dose depended limitation of ALP rise after CCl₄ administration. Although the dose of 100mg/kg for 21 days showed a significant limitation (p<0.05) of ALP rise (401±45.26) when compare to CCl₄ treated group but it did not match the efficacy of Liv. 52 treated group. However , in dose of 200 mg/kg for 21 days the *Rubia cordifolia* extract had much efficacy, in limiting the ALT rise after CCl₄ administration, to 245.1±21.25 IU/L, which was significant (p<0.01).

Table 3: Effect of Liv-52, *Rubia cordifolia* in their respective doses on carbon tetrachloride induced changes in total serum bilirubin (n=6)

TREATMENT (mg/kg)	Total bilirubin (IU/L) (mean±SE)
Normal saline (1ml)	0.25±0.95
CCl ₄ (1)	2.03±0.86 [^]
LIV.52 (1)	0.46±0.42 [^]
<i>Rubia cordifolia</i> (100)	0.93±0.53 [*]
<i>Rubia codifolia</i> (200)	0.83±0.13 [*]

£P< 0.05 as compare to ccl₄ treated group.

*p< 0.01 as compare to ccl₄ treated group.

[^]P< 0.001 as compare to normal saline treated group.

Total Bilirubin level in normal saline treated group was 0.25±0.95 IU/L. it was found to be significantly increased (p<0.001) with administration of CCl₄ to 2.03±0.86 IU/L. Pretreatment with known hepatoprotective preparation Liv.52 significantly (p<0.001) limited the rise in total bilirubin levels after CCl₄ administration to 0.46±0.42IU/L. With *Rubia cordifolia* there is dose depended limitation of Total Bilirubin rise after CCl₄ administration. Although the dose of 100mg/kg for 21 days showed a significant limitation (p<0.01) of Total Bilirubin rise (0.93±0.53) when compared to CCl₄ treated group but it did not match the efficacy of Liv. 52 treated group. However, in dose of 200 mg/kg for 21 days the *Rubia cordifolia* extract had much efficacy, in limiting the Total Bilirubin rise after CCl₄ administration, to 0.83±0.13 IU/L, which was significant (p<0.01).

Table 4: Effect of Liv-52, *Rubia cordifolia* in their respective doses on carbon tetrachloride induced changes in serum Albumin(mean± SE) (n=6)

TREATMENT (mg/kg)	Albumin (gm/dl) (mean±SE)
Normal saline (1ml)	4.6±0.82
CCl ₄ (1)	5.3±0.92
LIV.52 (1)	5.2±0.31
<i>Rubia cordifolia</i> (100)	5.4±1.14
<i>Rubia codifolia</i> (200)	5.04±0.94

There was no such difference in serum Albumin levels of any of groups and all the measurement recorded were in normal range. The Albumin levels varied from 4.2gm/dl to 5.4gm/dl showed no correlation. which was insignificant ($p>0.05$) also.

Effect on Histology

Histology of livers of normal saline treated group showed normal liver architecture. The hepatic cords and the sinusoids were well visible. (fig-1) Classical centrilobular necrosis was seen in the CCl₄ treated group. The hepatocytes around the central vein were necrosed with no distinguishable nuclei (fig-2).

Liv.52 treated group revealed vary mild signs of liver injury. Only difference from the normal saline treated group was the presence of inflammatory cells and constricted sinusoids indicating apperent hepatocyte swelling (fig-3).

Group treated with *Rubia cordifolia* with dose 100mg/kg showed feathery degeneration in the centrilobular area which is predominant histological feature, and necrosis was absent. However at dose 200mg/kg showed normal hepatic lobule architecture was seen with mild fatty changes and no necrosis was seen (fig.4-5).

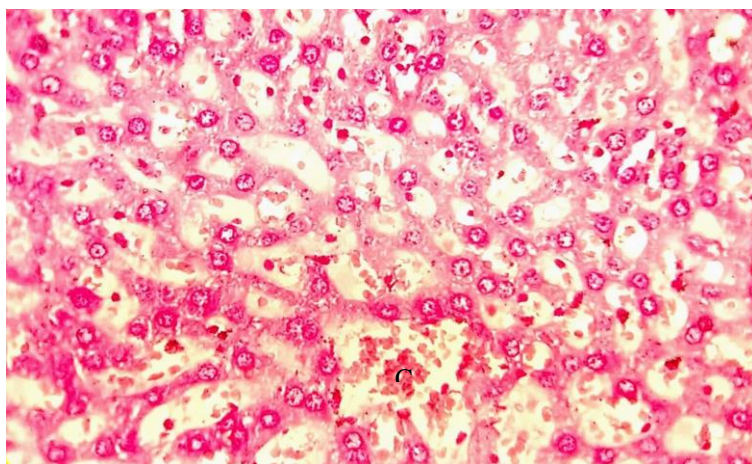


Figure 1: Microscopic feature of the liver normal saline treated group. Normal hepatic lobule architecture is seen. Hepatocytes and their nuclei are well visible. (H & E Stain x 400)

CV – Central vein

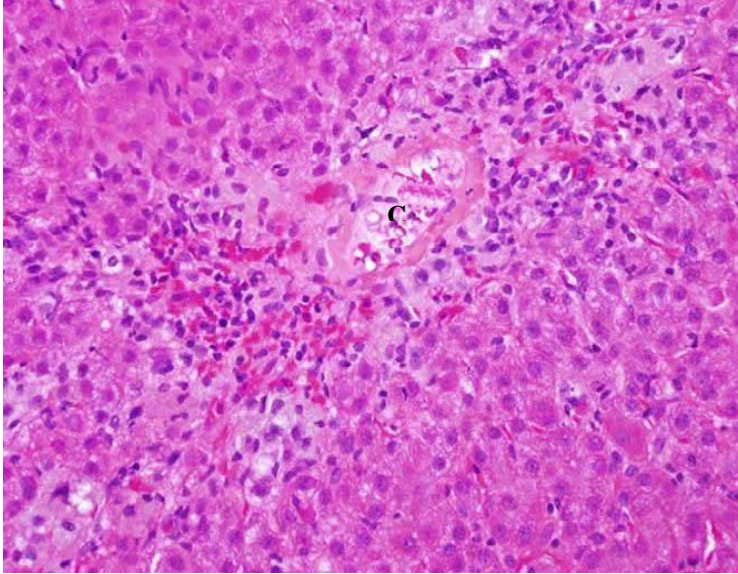


Figure 2: Microscopic feature of the liver CCl₄ treated group.
Extensive centrilobular necrosis is seen. Only cellular debris is seen and no hepatocytes with nuclei are discernible. (H & E Stain x 400)

CV – Central vein

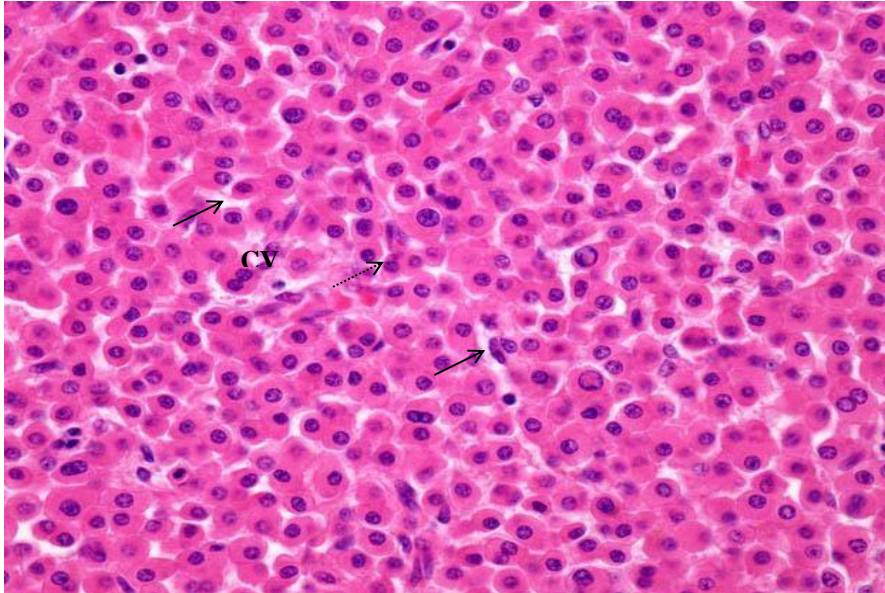


Figure 3: Microscopic feature of the liver Liv. 52 treated group.
Mild hepatocyte swelling is present as indicated by constructed sinusoids. Inflammatory cells are seen mostly around central vein. (H & E Stain x 400)

CV – Central vein

Solid Arrows : Sinusoids

Dotted Arrows : Inflammatory cells

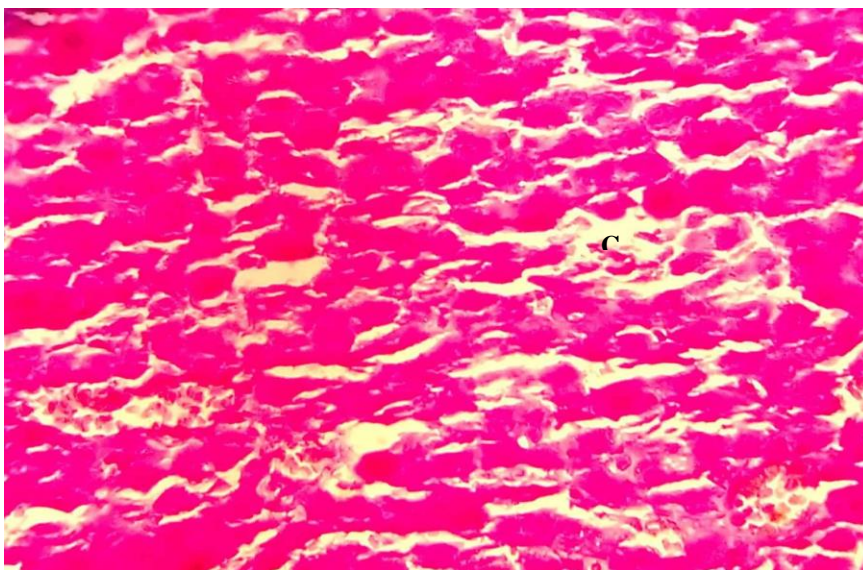


Figure 4: Microscopic feature of the liver of animals of group treated with *Rubia Cordifolia* (100 mg/kg) . Remarkable Feathery degeneration of haepatocyte is seen around the central vein. Further away from the central vein cell with fatty changes are seen. (H & E Stain x 400)

CV – Central vein

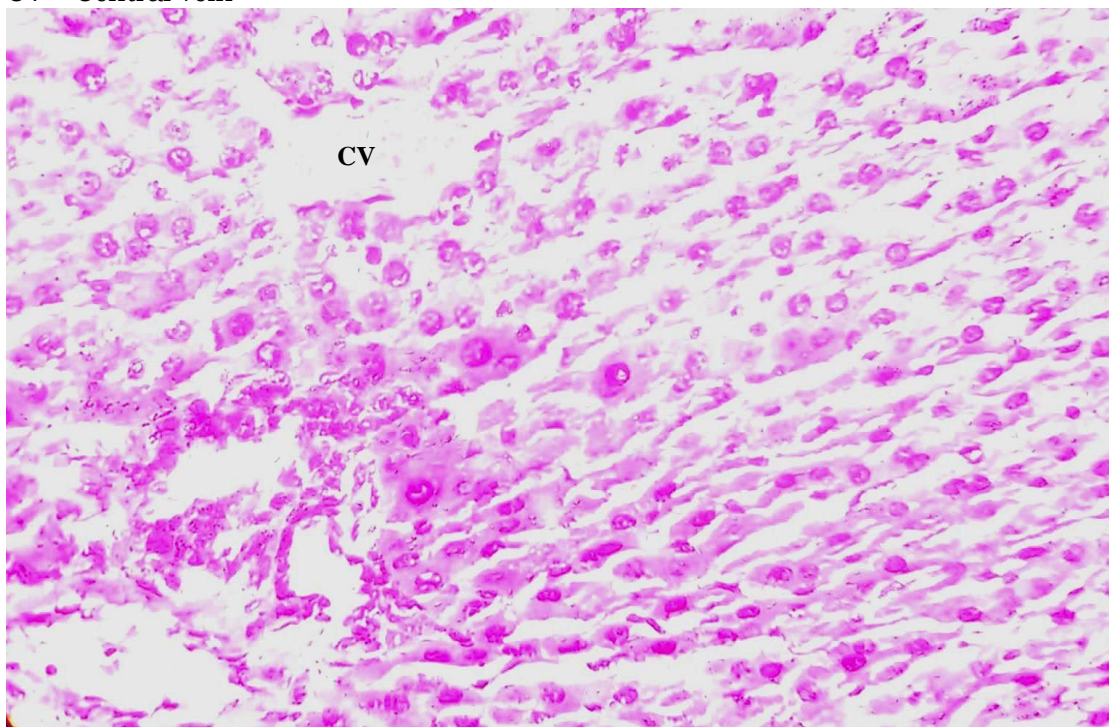


Figure 5: Microscopic feature of the liver of animals of group treated with *Rubia Cordifolia* (200 mg/kg) .

Miminal feathery degeneration around central vein. Cells with fatty changes are seen. Necrosis is not seen. (H & E Stain x 400)

CV – Central vein.

Discussion

Liver diseases are major problems throughout the world. Many environmental toxins cause liver diseases in human being. Despite new advances in hepatology, treatment of liver diseases caused by toxin has not been resolved as yet. Furthermore, despite the need for agents to protect the liver from damage, modern medicine lacks a reliable liver protective drug. [8] Therefore, there has been a considerable interest in the role of complementary and alternative medicines for the treatment of liver disease. [9] Recently so called healthy foods including functional food and dietary supplements are being increasingly used worldwide on part of complementary and supplementary medicine or self medication. [10]

The aqueous extract of *Rubia cordifolia* exhibited dose dependant hepatoprotection, both biochemically and histologically. Same dose for same duration also gave good protection from increasing ALP as compared to Liv.52 and slightly less protection from increasing Total Bilirubin as compare to Liv.52. No significant change was seen with albumin levels.

The hepatoprotective effect of *Rubia cordifolia* may be related to glutathione-mediated detoxification. Glutathione (g-glutamylcysteinyl glycine, GSH) is a sulfhydryl (SH) antioxidant, antitoxin, and enzyme cofactor. Glutathione is found mainly in the cell cytosol and other aqueous phases of the living system. [11] Glutathione exist in two form: the antioxidant – reduced glutathione tripeptide is conventionally called glutathione and abbreviated GSH; the oxidized form is a sulfur-sulfur linked compound, known as glutathione disulfide or GSSG. The GSSG/GSH ratio may be a sensitive indicator of oxidative stress. GSH has potent electron-donating capacity, as indicated by the high negative redox potential of the GSH/GSSH -redox couple ($E^{\circ} = -0.33\text{v}$). [12] Its high redox potential renders GSH both as a potent antioxidants and a convenient cofactor for enzymatic reactions that require readily available electron pairs. [13] Intracellular GSH status is a sensitive indicator of the cells overall health, and of its ability to resist toxic challenge. *Rubia cordifolia* and could also enhance the GSH in cells thereby may protect hepatic cells from toxic damages. [14]

Since formation of free radicals by cytochrome P450 after metabolism of CCl_4 has been implicated for lipid peroxidation mediated hepatocyte injury, the hepatoprotective mechanisms of aqueous extract of *Rubia cordifolia* could be an inhibitory effect on the microsomal enzymes (cytochrome P450) so that generation of free radicals is limited.

Like silymarin, *Rubia cordifolia* and *Withania somnifera* may also act as free radical scavengers and has antioxidant property as it may have an inhibitory effect on lipid peroxidation, Silymarin which is a well known hepatoprotective

agent. The hepatoprotective effects of silymarin are mainly attributable to its antioxidant and free radical scavenging properties. [15]

Hepatocytes have a practically unlimited capacity for proliferation, with full regeneration. Possible mechanism for regeneration of liver are changes in gene expression, an array of transcription factors (NF- κ B, STAT3, fos and jun) and various hepatic mitogenes which promote hepatocyte regeneration. Like silymarin, a proved hepatoprotective agent (acts through entry inside the nucleus and stimulate protein synthesis), may enhance *Rubia cordifolia* the activity of one or more of the above factors to stimulate the hepatic regeneration. [16]

Conclusion

Aqueous extract *Rubia cordifolia* is a potent hepatoprotective agent. Further studies with its hydroalcoholic extract can be expected to provide still better results. It is suggestible that the dosage form of *Rubia cordifolia* aqueous extract can be changed from the current *Rubia cordifolia* aqueous extract form, to some other form like hydroalcoholic extract or extraction of some specific component which is involved in hepatoprotective action. Also the safety profile of *Rubia cordifolia* aqueous extract has been very encouraging in this study. Direct human studies can be undertaken without any risk, which will help to determine the exact status of *Rubia cordifolia*. Further studies of extract of *Rubia cordifolia* could be extended for the isolation and structure determination of the hepatoprotective principle(s) and better and detailed explanations of mechanisms by which *Rubia cordifolia* acts.

References

1. Agrawal SS, Garg A, Agrawal S. Screening of *Phyllanthus niruri* Linn, and *Ricinus communis* Linn, on alcohol induced liver cell damage in non-hepatectomized and partially hepatectomized rats. Indian Journal of Pharmacology. 1986;18(4):211-214.
2. Ahmed B, Khan S, Masood MH, Siddique AH. Anti-hepatotoxic activity of cichotyboside, a sesquiterpene glycoside from the seeds of *Cichorium intybus*. J Asian Nat Prod Res. 2008 Mar-Apr; 10(3-4): 223-31.
3. Akahori A, Masui M, Kagawa K, Time course of biochemical and histological alterations following a single feeding of carbon tetrachloride to mice. Jpn J Exp Med 1983; 53: 199.
4. Alarcon de la Lastra AC, Martin MJ, Motilva V, Jimenez M, La Casa C, Lopez A. Gastroprotection induced by silymarin, the hepatoprotective principle of *Silybum marianum* in ischemia-reperfusion mucosal Injury: role of neutrophils. Planta Med. 1995 Apr; 61(2): 116-9.
5. Alcoholic liver disease. [Online]. 2005 Nov [cited 2008 Nov 10]; Available from: URL:<http://www.merck.com/mmDe/sec03/ch025/ch025a.html>
6. Anand KK, Chand D, Chandan BK. An evaluation of *Lawsonia alba* extract as hepatoprotective agent. Plant Med. 1992; 58: 22-5.
7. Anand KK, Gupta VN, Rangari V, Singh B, Chandan BK. Structure and hepatoprotective activity of biflavonoid from *Canarium manii*. Plant Med. 1992; 58: 493-4.

8. Ansari RA, Tripathi SC, Patnaik GK, Dhawan BN. Antihepatotoxic properties of picroliv and other fraction from rhizome of *Picrorhiza kurruoa*. *J Ethnopharmacol.* 1991; 34: 61-8.
9. Avadhoot Y, Rana AC. Hepatoprotective effect of *Vitex negundo* seeds against carbon tetrachloride induced liver damage. *Arch Pharmacol Res.* 1991; 14: 96-8.
10. Bacon BR. Cirrhosis and its complications. In: Fauci AS, Kasper DL, Longo DL, Braunwald E, Mauser SL, Jameson JL et al editors. *Harrison's principles of internal medicine.* 17th ed. New York: McGraw-Hill, Medical publishing division; 2008. p. 1972
11. Barker LF, Shulman NR, Murray R, (1996). "Transmission of serum hepatitis. 1970". *JAMA* 276 (10): 841-4.
12. Barston AD, Cerny EA, Tracy KM. Effect of administration of thioacetamide or testosterone on chromosome associated protein in rat liver and kidney. *Archives of Biochemistry and Biophysics.* 1965; 109: 36-40.
13. Benedek B, Kopp B. *Achillea millefolium* L. s.l. revisited: recent findings confirm the traditional use. *Wien Med Wochenschr.* 2007; 157(13-14): 312-4.
14. Bhise SB, Dias RJ. Monoethylglycinexylidide (MEGX) as a liver function test in cirrhosis. *Indian J Gastroenterol* 2007;26:167-9.
15. Bishayee A, Chatterhee MY. Carrot aqueous extract protection against hepatic oxidative stress and lipid peroxidation induced by acute carbon tetrachloride intoxication in mice. *Btoterapia.* 1993; 64: 261-5.
16. Bowman RC, Rand MJ. *Textbook of Pharmacology* 2nd ed. Oxford London. Blackwell Scientific Publications. 1982; 34-39.